

First Report of Two Cephalobidae Species (Nematoda: Cephalobomorpha) in South Korea

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ABSTRACT

Cephalobus aff. quinilineatus (Shavrov, 1968) Anderson and Hooper, 1970 and Eucephalobus hooperi Marinari-Palmisano, 1967 from the family Cephalobidae Filipjev, 1934 (Cephalobomorpha) are newly reported from South Korea. Cephalobus aff. quinilineatus is distinguished from other Cephalobus species by its high and rounded labial probolae and five lateral incisures, with three incisures extending to the tail terminus. Eucephalobus hooperi is distinguished from other Eucephalobus species by its three bifurcated labial probolae with pointed termini and by morphometric characters such as body and tail length and the corpus:isthmus ratio. In this study, the morphological characters and morphometrics of C. aff. quinilineatus and E. hooperi Korean population are described and illustrated based on optical and/or scanning electron microscopy.

Keywords: Cephalobidae, Cephalobus, Eucephalobus, new record, South Korea

INTRODUCTION

Members of the family Cephalobidae Filipjev, 1934 are bacterial feeders and are widely distributed in almost all terrestrial environments, including the dry soils of Antarctica and desert environments and tropical rainforests (Loof, 1971; Timm, 1971; Waceke et al., 2005; Nadler et al., 2006; Amirzadi et al., 2013). They are morphologically and taxonomically one of most problematic free-living nematode families (Andrássy, 2005) due to morphological and morphometric variation (such as body size, cephalic and labial probolae shape, nerve ring and excretory position and tail form) (Anderson, 1968; Anderson and Hooper, 1970, 1971; De Ley et al., 1993). The variability and similarity of morphological characters among Cephalobidae species obstruct species identification and phylogenetic studies for this group (Rashid et al., 1988; Boström, 1993; Abolafia and Peña-Santiago, 2002, 2005, 2009). Currently, 38 genera have been established in this family; however, only 29 can be accepted as valid (Andrássy, 2005). In Korea, 5 species have been reported (Eun et al., 2016; Kim et al., 2016; Kim et al., 2017a, 2017b, 2017c): Acrobeles ciliatus von Linstow, 1877; Acrobeloides nanus (de Man, 1880) Anderson, 1968; A. varius Kim, Kim and Park, 2017; Eucephalobus oxyuroides (de Man, 1876) Steiner, 1936; and Pseudacrobeles (Pseudacrobeles) curvatus Kim, Kim and Park, 2017.

During a survey of several overgrown fields, natural forest and plots of farmland, *C*. aff. *quinilineatus* (Shavrov, 1968) Anderson and Hooper, 1970 and *E. hooperi* Marinari-Palmisano, 1967 were collected and isolated from soil samples. Here we provide detailed descriptions of the morphological characters and morphometrics of the Korean isolates of *C*. aff. *quinilineatus* and *E. hooperi*.

MATERIALS AND METHODS

Nematode isolation

Soil samples were collected from the ground below pear trees (Gongdo-eup, Anseong-si, Gyeonggi-do, South Korea [GPS coordinates: 37°01′01.1″N, 127°10′23.4″E]), overgrown fields (Mechuri Island, Daenam-ro, Danwon-gu, Ansan-si, Gyeonggi-do, South Korea [GPS coordinates: 37°12′02.9″N, 126°32′24.6″E]), and natural forest areas

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(Geojejungang-ro, Dongbu-myeon, Geoje-si, Gyeong-sangnam-do, South Korea [GPS coordinates: 34°47′04.2″N, 128°37′30.3″E]). Nematode specimens were extracted by sieving and the Baermann funnel method (Baermann, 1917).

Fixation and morphological observations

For fixation, nematode specimens were transferred to 2 mL purified water in a 15 mL tube, to which was quickly added 4 mL of 80°C TAF (2% triethanolamine and 7% formaldehyde). The fixed nematodes were dehydrated in accordance with Seinhorst (1959) and mounted in pure glycerin on HS slides (Shirayama et al., 1993). Nematode morphological characters were observed under an optical microscope (BX-51; Olympus, Tokyo, Japan) equipped with differential interference contrast. Morphometric characters were measured using a CoolSnap Photometrics color CCD digital camera (MP5.0-RTV-R-CLR-10; Photometrics, Tucson, AZ, USA) and the program QCapture Pro 5 (QImaging, Surrey, Canada).

Scanning electron microscope (SEM)

For SEM imaging of *C*. aff. *quinilineatus*, the fixed specimens were transferred to an aqueous osmium tetroxide solution (4%) and kept at 4°C for 1 day for post-fixation. Fixed nematode specimens were dehydrated through a series of increasing ethanol concentrations (10–100% absolute ethanol, with 1 hour per stage). The samples were dried using a Hitachi HCP-2 critical point drier (Tokyo, Japan). Dried nematodes were mounted on a stub using copper/nickel tape and coated with gold/palladium using an Eiko IB-3 sputter-coater (Tokyo, Japan). Morphological characters of the nematode specimens were observed with a Zeiss Ultra Plus SEM (Oberkochen, Germany) at 15 kV under high-vacuum conditions.

SYSTEMATIC ACCOUNTS

Order Rhabditida Chitwood, 1993 Suborder Tylenchina Thorne, 1949 Infraorder Tylenchomorpha De Ley and Blaxter, 2002 Family Cephalobidae Filipjev, 1934 ^{1*}Genus *Cephalobus* Bastian, 1865

^{2*}Cephalobus aff. quinilineatus (Shavrov, 1968)
 Anderson and Hooper, 1970 (Table 1, Figs. 1, 2)

Chiloplacus quinilineatus: Shavrov, 1968: 137, fig. 1. Acrobeloides amurensis: Truskova, 1971: 434, fig. 1. Cephalobus quinilineatus: Anderson and Hooper, 1970: 468.

Material examined. 5♀♀ and 3♂♂, South Korea: Gyeonggi-do, Anseong-si, Gongdo-eup, Deokbongseowon-ro, 1 Jan 2017, extracted by sieving and the Baermann funnel method from soil below pear trees. Two specimens (slide Nos. NIBRIV0000812916 [female] and NIBRIV0000812917 [male]) are deposited at the National Institute of Biological Resources, Republic of Korea and six specimens (slide Nos. 01010202002-01010202005 [females] and 01010202007 and 01010202008 [males]) are deposited in the Animal Phylogenomics Laboratory, Ewha Womans University, Republic of Korea.

Measurements. See Table 1.

Description. Adult: Female body cylindrical, length 865.8– 962.4 µm; ventrally curved after fixation, C-shaped. Male body cylindrical, length 808.1-908.9 µm; posterior region more curved than female, J-shaped. Cuticle annulated; annuli 2.4-3.1 μm wide and 1.0-1.7 μm thick at mid-body. Lateral field occupying 20.0-24.3% of width of body at mid-body in female, 32.3-35.1% of body diameter in male. Lateral incisures varying in number along body length: two incisures at procorpus region, branching off from posterior corpus into five incisures, two incisures fading out at phasmid region, three incisures extending to tail terminus. Head region continuous with neck. Lip region 9.8-11.9 µm in diameter, with triradiate symmetry with 6+4 papillae. Six lips grouped in pairs; deep U-shaped primary axils; shallow V-shaped secondary axils; lacking guarding processes, without tine. Cephalic probolae absent. Three plate-like, high and rounded labial probolae present. Transversal, ovalshaped amphidial apertures present. Stoma cephaloboid, 1.4-1.6 times the lip region diameter in length; cheilorhabdions oval shaped. Pharyngeal corpus cylindrical, 2.9-4.2 times isthmus length. Isthmus narrower than corpus, distinctly demarcated from metacorpus. Basal bulb ovalshaped with well developed valves; 1.1-1.3 times as long as its width. Cardia conoid, surrounded by intestinal tissue. Nerve ring position varying from posterior corpus to anterior isthmus, at 65.7-72.9% of pharynx length. Excretory pore position similar to nerve ring, at 67.8-79.5% of pharynx length. Deirids positioned in lateral field at level of isthmus or isthmus-basal bulb junction, at 75.7-87.1% of total neck length.

Female: Reproductive system monodelphic-prodelphic. Vulva lips slightly protruding or not. Vagina short, length 0.3–0.6 times the body diameter. Post-uterine sac 0.8–2.1 times the body width. Uterus length 2.9–4.0 times the body diameter. Spermatheca inconspicuous, 1.3–2.1 times the body width. Oviduct short. Ovary with or without double flexure posterior to vulva. Rectum length 1.2–1.5 times the

 Table 1.
 Morphometrics of Cephalobus aff. quinilineatus and Eucephalobus hooperi

متملروتوطي	Cephalobus aff. quinilineatus	uinilineatus	Eucephalobus hooperi	peri
כומומרופוט	₽, n=5	σ^2 , n=3	$^{\circ}$, n=4 (Ansan)	4, n=1 (Geoje)
	902.9±44.4 (865.8–968.4)	848.6±53.3 (808.1–908.9)	416.5±48.9 (366.3–483.1)	452.6
ھ	21.6 ± 1.7 (19.3–24.0)	$24.6\pm0.9(23.5-25.3)$	21.8 ± 2.2 (19.5–23.9)	21.2
q	4.5 ± 0.3 $(4.2-4.8)$	$4.5\pm0.3(4.3-4.9)$		3.5
v	$20.2\pm0.5(19.4-20.6)$	$16.7\pm0.8(16.2-17.6)$	$9.1\pm1.1(7.5-10.1)$	10.1
Ú		$1.8\pm0.1(1.7-1.9)$	4.1 ± 0.2 (3.8–4.4)	3.5
>	$66.4 \pm 1.9 (64.8 - 69.4)$		67.5 ± 6.2 ($63.7-76.7$)	65.5
GorT	2.2		_	25.7
Body diameter	42.0 ± 2.7 (38.6–44.9)		_	21.3
Pharynx length				129.2
Tail length	$44.7 \pm 2.8 (42.3 - 47.8)$	$50.7 \pm 1.3 (49.3 - 51.6)$	$46.9\pm11.7~(39.9-64.4)$	45
Anal body diameter	$23.3\pm1.7~(20.5-24.9)$	$27.7 \pm 1.9 (26.4 - 29.9)$	$11.5\pm2.2\ (10.1-14.7)$	12.7
Lip region diameter	$11.0\pm0.8~(9.8-11.9)$	$10.4\pm0.2(10.1\!-\!10.6)$		5.7
Stoma	$15.9\pm0.4~(15.5-16.3)$			10.8
Stoma diameter	$6.6\pm0.6(5.7-7.0)$		$4.3\pm0.6(3.7-5.1)$	4
Stoma/lip region diameter	$1.5\pm0.1(1.4{-}1.6)$		$1.8\pm0.2(1.6-2.0)$	1.9
Stoma/stoma diameter	$2.4\pm0.2(2.2-2.8)$		$2.5\pm0.1(2.3-2.6)$	2.7
Corpus	$119.1 \pm 7.6 (109.7 - 130.0)$		$75.5\pm17.5(61.1-101.0)$	73.9
Isthmus	$32.7 \pm 4.2 (26.0 - 37.1)$	$34.7 \pm 1.4 (33.7 - 36.3)$		24.4
Basal bulb	25.3±3.3 (20.6–28.7)	$22.7 \pm 0.5 (22.2 - 23.1)$		15.6
Basal bulb diameter	$21.5\pm1.3(19.6-22.9)$	$18.8\pm0.3(18./-19.1)$		12.5
Basal bulb length/diameter	0.1	$1.2\pm0.0(1.2-1.2)$	$1.3\pm0.0(1.3-1.4)$	1.2
Corpus:Istnmus ratio	3.7±0.4 (3.2-4.2)	3.1±0.3(2.9−3.4) 124 E±2 0(122 € 141 E)	2.9±0.5 (2.4–3.6)	ς, c ₀
Exercise to all the part and	130.1±0.3 (131.4-140.1)			05.1
Excretory pore to ant. end Deirid to ant end	0.7		04.9±14.0 (/4./−100.0) 04.4+13.3 (83.3−113.7)	90.7 97.5
Nerve ring position (% pharvnx)			62.5+3.9 (59.1–67.9)	64.3
Excretory pore position (% pharvnx)	73.6±4.3 (67.8–79.5)		$63.9 \pm 0.5 (63.2 - 64.5)$	67.1
Deirid position (% pharynx)	$82.7 \pm 3.2 (79.0 - 86.4)$			75.5
Vulva from ant. end	599.3±22.6 (572.7–630.1)	, I		296.6
Vulva to anus	258.5±23.3 (229.3–283.4)	ı	110.4 ± 24.6 (89.9–145.2)	107.4
Vulva to anus/tail length	$5.8\pm0.3(5.4-6.2)$	ı	$2.4\pm0.2(2.3-2.7)$	2.4
Reproductive tract length	$424.6\pm39.5\ (399.5-491.4)$	$408.2\pm80.4(317.4-470.3)$	133.3 ± 42.5 (105.4–196.4)	116.3
Vagina	$19.7 \pm 4.0 (13.8 - 24.3)$	I	7.3 ± 1.9 (5.7–10.1)	2.6
Post-uterine sac	$72.6\pm20.8(37.1-88.9)$	I	$24.0\pm2.3(21.4-26.9)$	21.7
Uterus	144.2±14.0 (128.3–156.1)	ı	$43.1 \pm 9.6 (37.8 - 57.6)$	45.3
Spermatneca	65.6±16.5 (52.7–93.2)	I	23.8±7.9 (16.1–33.6)	34.6
Oviduct	$1/.0\pm/.0$ (9.2–26.7)	1	$8.0\pm 2.3(5.6-11.0)$	10.1
Ovary	331.3±33.0 (315.9-392.5) 0 5±0 1 (0 3-0 6)	1	105.3±40.1 (82.4-165.3)	90
Vagilla/body diallieter	0.0±0.1 1 7±0 = (0 0 0 1)	1	0.4±0.0 (0.3-0.4)	c
rost-dreillie sac/ body dialiterei Herris/hody diameter		l 1		2.1
Spermathera/hody diameter	1 6+0 3 (1 3-2 1)	1		1.1
Oviduct/hody diameter	2.0	ı	$0.4 \pm 0.1 (0.3 - 0.5)$	0.5
Ovary/body diameter	0.9	ı	_	4.2
Spicules		$50.9\pm3.4(47.0-53.6)$, 1	ı
Spicules/body width	1 1	$1.5\pm0.1(1.4-1.6)$	1 1	1 1
	1	23.0 ± 3.0 (22.3 – 23.6)	1	

Table 1. Continued

Gubernaculum/body width Gubernaculum/spicules	₽, n=5 _			
Gubernaculum/body width Gubernaculum/spicules	ı	o², n=3	♀, n=4 (Ansan)	♀, n=1 (Geoje)
Gubernaculum/spicules		0.7±0.0(0.7-0.8)	ı	ı
	I	$0.5\pm0.1(0.4-0.6)$	ı	1
Rectum 30	$30.8\pm1.8(29.0-33.8)$, 1	$15.7 \pm 3.9 (12.6 - 21.4)$	16.6
Rectum/anal body diameter	$1.3\pm0.1(1.2-1.5)$	1	$1.4 \pm 0.1 (1.2 - 1.5)$	1.3
Anus to phasmid 29	29.9 ± 3.6 (26.4–35.4)	$33.4 \pm 2.6 (31.6 - 36.5)$	$13.5\pm1.3(12.8-15.4)$	13.2
(% tail)	$66.8\pm6.6(61.2-74.3)$	$65.9\pm4.5(61.6-70.6)$	$30.0\pm6.9(20.2-36.6)$	29.3
Lateral field width	$9.4\pm1.0(7.7-10.4)$	$11.8 \pm 0.7 (11.2 - 12.5)$	$3.4\pm0.5(2.8-4.0)$	4.5
Lateral field width/body diameter (%)	$22.3 \pm 1.7 (20.0 - 24.3)$	$34.1 \pm 1.6 (32.3 - 35.1)$	$17.6 \pm 1.6 (16.0 - 19.4)$	21.1
Cuticle thickness	$1.3\pm0.3(1.0-1.6)$	$1.4\pm0.3(1.1-1.7)$	1.2 ± 0.1 $(1.0-1.2)$	1.2
Annuli width	$2.9\pm0.3(2.6-3.1)$	$2.7\pm0.3(2.4-3.0)$	$1.5\pm0.2\ (1.2-1.8)$	1.6
Mucro	1	I	$4.9\pm5.1(1.3-8.5)$	ı

anal body diameter. Tail conoid, with rounded terminus. Phasmids located behind middle of tail, at 61.2-74.3% of tail length.

Male: Genital system monorchic. Testis reflexed ventrad anteriorly. Spicules curved ventrad, 47.0-53.6 µm long; manubrium rounded or conoid; calamus wider than manubrium; without hump or velum; with one longitudinal incisure. Gubernaculum curved ventrad, sigmoid. Two pairs of pre-cloacal subventral papillae present. Post-cloacal genital papillae in five pairs: one subventral and one lateral at midtail; one lateral, one ventral and one subdorsal at tail terminus region. Tail conoid, blunt terminus. Phasmid opening at 61.6-70.6% of tail length.

Distribution. India, Russia, South Korea.

Habitat. Soil sample from a pear farm.

Remarks. Morphological characters and measurements of the specimens described in this study generally agree with original descriptions of C. quinilineatus (Shavrov, 1968) Anderson and Hooper, 1970, except for female body length (865.8-968.4 vs. 642-712 µm) and labial probolae shape (rounded vs. bifurcated) (Shavrov, 1968). Although female body length and labial probolae shape differ between the present specimens and the original description of C. quinilineatus, this is not sufficient evidence for a different species diagnosis, due to the varying degrees of bifurcation and rounded labial probolae of the synonymized A. amurensis (Shavrov, 1968; Truskova, 1971). Additionally, the original description of C. quinilineatus did not include the male and omitted many key characters needed for identification to species level, such as descriptions of the lateral field at the tail region, post-uterine sac, spermatheca and ratio of corpus to isthmus length (Shavrov, 1968). Therefore, it is impossible to be certain about the identity of the specimens. Due to close similarity of the specimens with C. quinilineatus, we consider the specimens to be C. aff. quinilineatus. This species is reported for the first time from South Korea.

Genus Eucephalobus Steiner, 1936

Material examined. 4♀♀, South Korea: Gyeonggi-do: Ansan-si, Danwon-gu, Daenam-ro, Mechuri Island, 22 Sep 2016; 1♀, Gyeongsangnam-do: Geoje-si, Dongbu-myeon, Geojejungang-ro, 5 Jul 2017, extracted by sieving and the Baermann funnel method from overgrown field and natural forest soil, respectively. The two specimens (slide Nos. NI-BRIV0000812922 and NIBRIV0000812923) are deposited

Korean name: 1*후퍼유럽두옆선충 (신칭)

^{1*}Eucephalobus hooperi Marinari-Palmisano, 1967 (Table 1, Fig. 3)

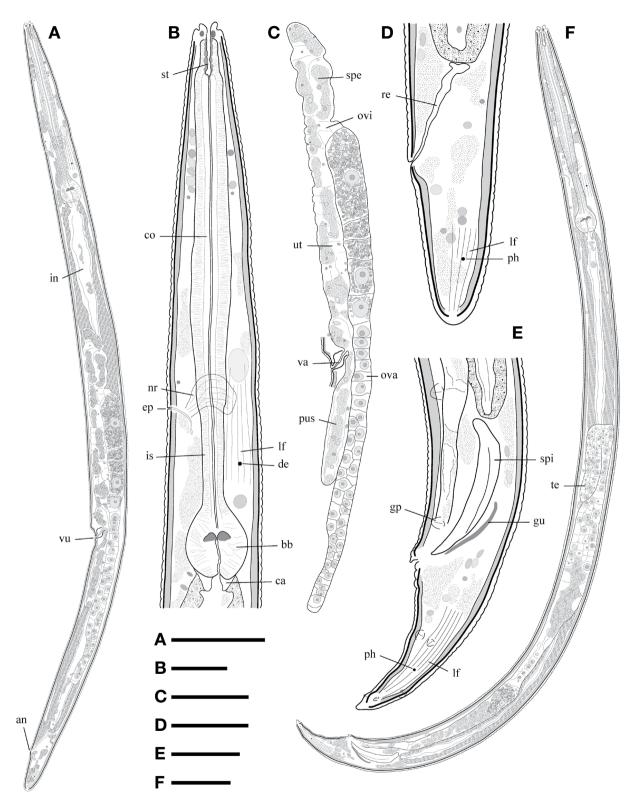


Fig. 1. *Cephalobus* aff. *quinilineatus* (Shavrov, 1968) Anderson and Hooper, 1970. A, Entire female; B, Female neck region; C, Female reproductive system; D, Female posterior region; E, Male posterior region; F, Entire male. an, anus; bb, basal bulb; ca, cardia; co, corpus; de, deirid; ep, excretory pore; gp, genital papilla; gu, gubernaculum; in, intestine; is, isthmus; lf, lateral field; nr, nerve ring; ova, ovary; ovi, oviduct; ph, phasmid; pus, post-uterine sac; re, rectum; spe, spermatheca; spi, spicule; st, stoma; te, testis; ut, uterus; va, vagina; vu, vulva. Scale bars: A=100 μm, B, D, E=20 μm, C, F=50 μm.

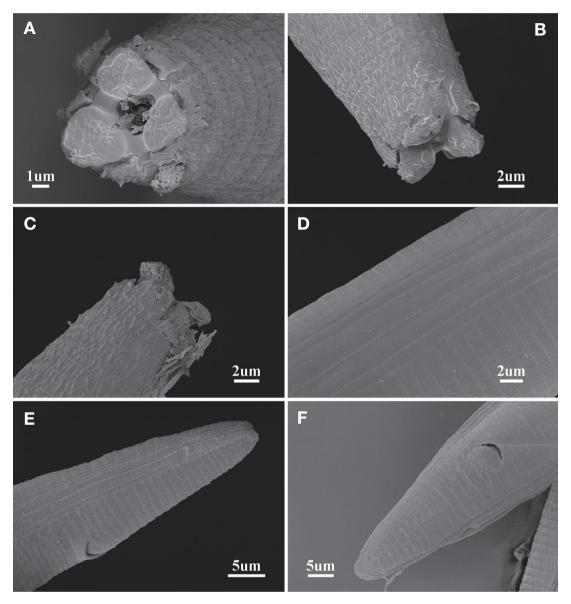


Fig. 2. Cephalobus aff. quinilineatus (Shavrov, 1968) Anderson and Hooper, 1970. A, Lip region, en face view; B, Lip region, lateral view; C, Lip region, ventral view; D, Lateral field; E, Tail, lateral view; F, Tail, ventral view. Scale bars: A=1 μm, B-D=2 μm, E, F=5 μm.

at the National Institute of Biological Resources, Republic of Korea, and three specimens (slide Nos. 01010103001, 01010103002 and 01010103007) are deposited in the Animal Phylogenomics Laboratory, Ewha Womans University, Republic of Korea

Measurements. See Table 1.

Description. Female: Body cylindrical, length 366.3–483.1 μm, usually ventrally curved after fixation. Cuticle annulated; annuli 1.2–1.8 μm wide and 1.0–1.2 μm thick at mid-body. Lateral field occupying 16.0–21.1% of width of body at mid-body. Three incisures in lateral field, fading

out at phasmid region. Head region continuous with neck. Six lips conoid, bearing 6+4 papillae. Transversal, oval-shaped amphidial apertures present. Three bifurcate labial probolae connected basally by tangential ridges, with seta-like or pointed termini. Stoma cephaloboid, length 1.6-2.0 times the lip region diameter. Cheilorhabdions oval-or bar-shaped. Small dorsal denticle sometimes present on metastom. Pharyngeal corpus cylindrical, 2.4-3.6 times isthmus length. Isthmus narrower than corpus, distinctly demarcated from metacorpus. Basal bulb oval-shaped with well developed valves; 1.2-1.4 times as long as its width.

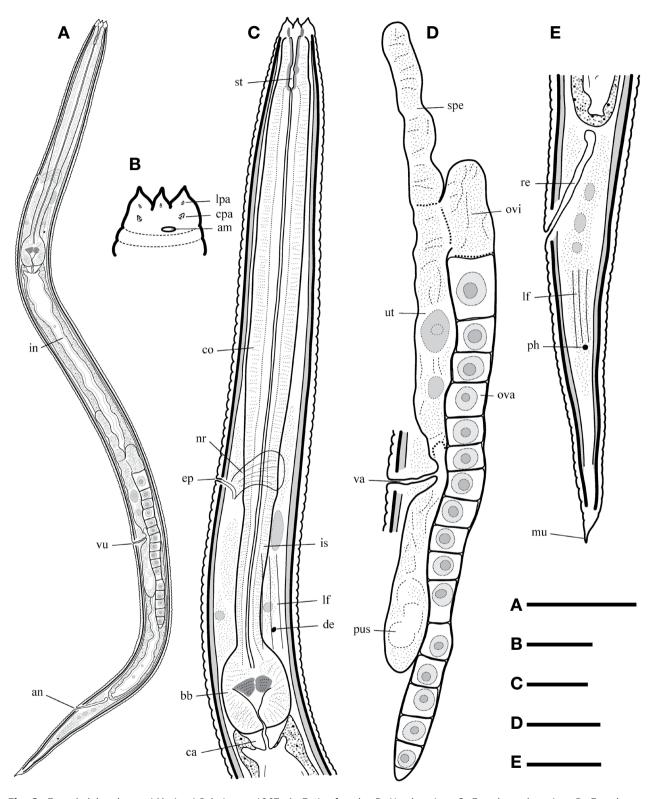


Fig. 3. *Eucephalobus hooperi* Marinari-Palmisano, 1967. A, Entire female; B, Head region; C, Female neck region; D, Female reproductive system; E, Female posterior region. am, amphid; an, anus; bb, basal bulb; ca, cardia; co, corpus; cpa, cephalic papilla; de, deirid; ep, excretory pore; in, intestine; is, isthmus; If, lateral field; lpa, labial papilla; mu, mucro; nr, nerve ring; ova, ovary; ovi, oviduct; ph, phasmid; pus, post-uterine sac; re, rectum; spe, spermatheca; st, stoma; ut, uterus; va, vagina; vu, vulva. Scale bars: $A = 50 \mu m$, $B = 5 \mu m$, $C - E = 10 \mu m$.

Cardia conoid, surrounded by intestinal tissue. Nerve ring located at posterior corpus or corpus-isthmus junction, at 59.1-67.9% of pharynx length. Excretory pore position at posterior corpus or corpus-isthmus junction, at 63.2-67.1% of pharynx length. Position of deirids in lateral field at isthmus or corpus-isthmus junction, at 68.7-75.5\% of total neck length. Reproductive system monodelphic-prodelphic. Vulva lips not protruding. Vagina length 0.3-0.4 times the body diameter. Post-uterine sac 1.0-1.6 times the body width. Uterus 2.1-2.3 body diameters long. Spermatheca 0.9-2.0 times the body width. Oviduct short. Ovary straight, with a single row of oocytes. Rectum length 1.2-1.5 times the anal body diameter. Tail elongated-conoid, with pointed terminus. Spike-shaped mucro sometimes present. Phasmids located anterior to middle of tail, at 20.2-36.6% of tail length. Male: Unknown.

Distribution. India, Italy, Kenya, Malaysia, South Korea, Spain.

Habitat. Soil sample from overgrown field and forest soil. **Remarks.** Morphological characters of the specimen described in this study generally agree with other previous studies (Marinari-Palmisano, 1967; Boström, 1990, 1993; Abolafia and Peña-Santiago, 2002). However, the Korean population differs from the Italian populations in the ratio of corpus to isthmus length (2.4–3.6 vs 5–6) (Marinari-Palmisano, 1967). Compared to the Malaysian population, it differs in esophagus length relative to total body length (b=2.9–3.5 vs. 3.6–3.8) and length of post-uterine sac relative to body width (1.0–1.6 vs. 0.7–0.8) (Boström, 1993). *Eucephalobus hooperi* is reported for the first time from South Korea.

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REFERENCES

- Abolafia J, Peña-Santiago R, 2002. Nematodos del orden Rhabditida de Andalucía Oriental. El género *Eucephalobus* Steiner, 1936. Graellsia, 58:59-78.
- Abolafia J, Peña-Santiago R, 2005. Nematodes of the order Rhabditida from Andalucía Oriental: *Pseudacrobeles elon*-

- *gatus* (de Man, 1880) comb. n. Nematology, 7:917-926. https://doi.org/10.1163/156854105776186415
- Abolafia J, Peña-Santiago R, 2009. Nematodes of the order Rhabditida from Andalucía Oriental, Spain. The genus *Cephalobus* Bastian, 1865 with description of *C. harpagonis* sp. n. and key to species. Nematology, 11:485-508. https://doi.org/10.1163/138855409X12465362560359
- Amirzadi N, Shokoohi E, Abolafia J, 2013. Description of nine species of the family Cephalobidae (Nematoda, Rhabditida) and morphometric analysis in the genus *Acrobeles* von Linstow, 1877. Acta Zoologica Bulgarica, 65:3-26.
- Anderson RV, 1968. Variation in taxonomic characters of a species of *Acrobeloides* (Cobb, 1924) Steiner and Buhrer, 1933. Canadian Journal of Zoology, 46:309-320. https://doi.org/10.1139/z68-048
- Anderson RV, Hooper DJ, 1970. A neotype for *Cephalobus persegnis* Bastian, 1865, redescription of the species, and observations on variability in taxonomic characters. Canadian Journal of Zoology, 48:457-469. https://doi.org/10.1139/z70-078
- Anderson RV, Hooper DJ, 1971. A neotype for *Eucephalobus striatus* (Bastian, 1865) Thorne, 1937 (Nematoda) and redescription of the species from topotypes and their progeny. Canadian Journal of Zoology, 49:451-459. https://dx.doi.org/10.1139/z71-070
- Andrássy I, 2005. Free-living nematodes of Hungary: Nematoda errantia. Volume I. Hungarian Natural History Museum, Budapest, pp. 1-518.
- Baermann G, 1917. Eine einfache methode zur auffindung von ankylostomum (Nematoden) larven in erdproben. Geneeskundig Tijdschrift voor Nederlandsch-Indië, 57:131-137.
- Boström S, 1990. Some species of Cephalobidae (Nematoda: Rhabditida) from highland Kenya. Journal of African Zoology, 104:127-134.
- Boström S, 1993. Some cephalobids from Ireland and Malaysia (Nematoda: Rhabditida). Afro-Asian Journal of Nematology, 3:128-134.
- De Ley P, Siddiqi MR, Boström S, 1993. A revision of the genus *Pseudacrobeles* Steiner, 1938 (Nematoda: Cephalobidae). Part 1. Subgenus *Pseudacrobeles* grad. n. Fundamental and Applied Nematology, 16:219-238.
- Eun G, Ha J, Kang H, Kim Y, Choi I, Kim D, 2016. First record *Acrobeles ciliatus* (Rhabditida) and *Plectus parietinus* (Plectida) from South Korea. Journal of Species Research, 5:318-323. https://doi.org/10.12651/JSR.2016.5.3.318
- Kim J, Kim T, Park J-K, 2017a. Description of *Pseudacrobeles* (*Pseudacrobeles*) *curvatus* sp. n. (Cephalobidae: Rhabditida) in South Korea. Journal of Nematology, 49:162-167. https://doi.org/10.21307/jofnem-2017-061
- Kim T, Bae YJ, Park J-K, 2017b. First record of *Eucephalobus* oxyuroides (Nematoda: Rhabditida: Cephalobidae) from South Korea. Animal Systematics, Evolution and Diversity, 33:256-261. https://doi.org/10.5635/ASED.2017.33.4.027
- Kim T, Kim J, Bae YJ, Park J-K, 2016. First record of *Acrobeloides nanus* (Cephalobidae: Rhabditida: Nematoda) from

- Korea. Animal Systematics, Evolution and Diversity, 32: 258-265. https://doi.org/10.5635/ASED.2016.32.4.035
- Kim T, Kim J, Park J-K, 2017c. *Acrobeloides varius* sp. n. (Rhabditida: Cephalobidae) from South Korea. Nematology, 19: 489-496. https://doi.org/10.1163/15685411-00003064
- Loof PAA, 1971. Freeliving and plant parasitic nematodes from Spitzbergen, collected by Mr. H. van Rossen. Mededelingen Landbouwhogeschool Wageningen, 71:1-86.
- Marinari-Palmisano A, 1967. Contributo alla conoscenza di alcuni nematodi dei generi *Rhabditoides*, *Eucephalobus*, *Het*erocephalobus. Redia, 50:289-308.
- Nadler SA, De Ley P, Mundo-Ocampo M, Smythe AB, Stock SP, Bumbarger D, Adams BJ, De Ley IT, Holovachov O, Baldwin JG, 2006. Phylogeny of Cephalobina (Nematoda): molecular evidence for recurrent evolution of probolae and incongruence with traditional classifications. Molecular Phylogenetics and Evolution, 40:696-711. https://doi.org/10.1016/j.ympev.2006.04.005
- Rashid F, Geraert E, Coomans A, Suatmadji W, 1988. Cephalobidae from the Krakatau region (Nematoda: Rhabditida). Nematologica, 34:125-143. https://doi.org/10.1163/002825 988X00224
- Seinhorst JW, 1959. A rapid method for the transfer of nema-

- todes from fixative to anhydrous glycerin. Nematologica, 4:67-69. https://doi.org/10.1163/187529259X00381
- Shavrov GN, 1968. New species of plant nematodes of the subfamily Acrobelinae Thorne, 1937. Soobscenija Dal'nevostocnogo Filiala Akademii Nauk SSSR, 26:137-140 (in Russian).
- Shirayama Y, Kaku T, Higgins RP, 1993. Double-sided microscopic observation of meiofauna using an HS-slide. Benthos Research, 44:41-44.
- Timm RW, 1971. Antarctic soil and freshwater nematodes from the McMurdo Sound region. Proceedings of the Helminthological Society of Washington, 38:42-52.
- Truskova GM, 1971. A new species of *Acrobeloides* (Nematoda, Cephalobidae). Zoologicheskii Zhurnal, 50:434-436 (in Russian).
- Waceke JW, Bumbarger DJ, Mundo-Ocampo M, Subbotin SA, Baldwin JG, 2005. *Zeldia spannata* sp. n. (Nematoda: Cephalobidae) from the Mojave Desert, California. Journal of Nematode Morphology and Systematics, 8:57-67.

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